Probing the Kinetics of Single Molecule Protein Folding

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ABSTRACT We propose an approach to integrate the theory, simulations, and experiments in protein-folding kinetics. This is realized by measuring the mean and high-order moments of the first-passage time and its associated distribution. The full kinetics is revealed in the current theoretical framework through these measurements. In the experiments, information about the statistical properties of first-passage times can be obtained from the kinetic folding trajectories of single molecule experiments (for example, fluorescence). Theoretical/simulation and experimental approaches can be directly related. We study in particular the temperature-varying kinetics to probe the underlying structure of the folding energy landscape. At high temperatures, exponential kinetics is observed; there are multiple parallel kinetic paths leading to the native state. At intermediate temperatures, nonexponential kinetics appears, revealing the nature of the distribution of local traps on the landscape and, as a result, discrete kinetic paths emerge. At very low temperatures, exponential kinetics is again observed; the dynamics on the underlying landscape is dominated by a single barrier. The ratio between first-passage-time moments is proposed to be a good variable to quantitatively probe these kinetic changes. The temperature-dependent kinetics is consistent with the strange kinetics found in folding dynamics experiments. The potential applications of the current results to single-molecule protein folding are discussed.

INTRODUCTION

Protein folding is a central issue of modern science. It is fundamental for understanding biological structure and function, and it is also connected to many technological applications such as drug design. The crucial question regarding the folding mechanism is how the many possible conformational degrees of freedom converge to the unique native basin. Looking at details of the microscopic kinetics and figuring out the dominant pathways are important, but this approach is sometimes misleading and misses the global picture (Levinthal,1969; Baldwin, 1995; Laurents et al., 1988). The possibility that multiple paths leading to folding may exist has led to an alternative statistical approach to this problem (Bryngelson et al., 1995; Wolynes et al., 1995; Chan and Dill, 1994; Abkevich et al., 1994; Wang et al, 1996).

The resulting energy-landscape theory looks at a coarse-grained-level picture of folding and tries to uncover the general principles governing its mechanisms. In general, the energy landscape is rough due to the many different possible competing interactions among the amino acid residues. On the other hand, the landscape of real proteins has evolved toward a funnel, shaped with a gradient leading toward the native folded basin. There are in general many possible paths leading to folding. A small number of discrete pathways emerge only when the landscape becomes rough and local traps become significant (Wang et al., 1996). The predictions of the funneled-landscape picture of folding have been

confirmed and verified by many lattice and off-lattice simulations (Chan and Dill, 1994; Abkevich et al., 1994; Klimov and Thirumalai, 1998; Socci and Onuchic, 1994, 1995; Chahine et al., 2002; Kaya and Chan, 2000, 2002, 2003a,b,c; Boczko and Brooks, 1995). They are also in good qualitative and quantitative agreement with protein-folding thermodynamic and kinetic experiments (Itzhaki et al., 1995; Huang and Oas, 1995; Eaton et al., 1997, 2000; Garcia-Mira et al., 2002; Sabelko et al., 1999; Nguyen et al., 2003, Kuhlman et al., 1998).

The landscape theory of folding defines a temperature where the native protein structure becomes stable, $T_{\rm f}$. It also defines a glassy temperature, $T_{\rm g}$, where the landscape becomes rough compared with the bias toward a native folded state, and trapping into local minimum occurs. The ratio $T_{\rm f}/T_{\rm g}$ defines how good a folder a protein is, i.e., the energy gap should be larger than the underlying roughness of the landscape (Goldstein et al., 1992; Chan and Dill, 1994; Chan et al., 2004; Abkevich et al., 1994; Klimov and Thirumalai, 1998). The cooperativity in the physical interactions can influence the ratio of $T_{\rm f}/T_{\rm g}$ significantly (Chan et al., 2004).

There have been theoretical studies on average measurable variables of the folding kinetics (mean first-passage times, MFPT, or average rates) (Bryngelson and Wolynes, 1989; Saven et al., 1994; Wang et al., 1996; Shakhnovich et al., 1989; Gutin et al., 1996; Seno et al., 1998; Cieplak et al., 1999; Klimov and Thirumalai, 1998; Socci et al., 1996). Although the average-value description of the kinetics is important, it can sometimes miss important features of folding dynamics that are crucial for uncovering the funda-

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mental mechanisms. For different sequence compositions or external conditions (physical properties, temperatures, and solvent conditions) the progress of folding can probe different parts of the energy landscape and detect different local barriers. Therefore many possible timescales may coexist, and the kinetics can become nonexponential or multiexponential. In bulk measurements, it is often difficult to distinguish whether the observed nonexponential kinetics is intrinsic or due to the inhomogeneous distribution of single exponential processes. With recent technological advances, however, single-molecule detections have become possible (Moerner, 1996; Lu et al., 1998; Deniz et al., 2000). Single molecules are sensitive probes to the local environments and therefore an ideal tool to understand the structures of the energy landscape of the proteins (Wang and Wolynes, 1995, 1999; Onuchic et al., 1999; Wang, 2003a,b; Schenter et al., 1999). Lately, a number of remarkable initial singlemolecule folding experiments have been undertaken (Zhuang et al., 2000, 2002; Jia et al., 1999; Deniz et al., 2000; Schuler et al., 2002; Lipman et al., 2003) but their interpretation is nontrivial. Statistical fluctuations are intrinsic to single molecules, since they are not weighted down by the number of the molecules as they are in the bulk, and they can be directly measured. Single-molecule data are essentially sequences of on-and-off spikes as a function of time. Therefore, to determine whether they are associated with exponential decay is a challenging task.

In this article, we investigate the kinetics of folding based on the moments or fluctuations of the FPTs of folding and dynamic collapse, with respect to temperature and physical properties such as hydrophobicity. From these studies, we are able to determine the way in which the exponential or nonexponential nature of the folding kinetics will emerge as a function of these conditions. Furthermore, we obtain information about the dependence of the distribution of folding times upon various conditions. This gives the full dynamic picture of the folding instead of just the MFPT. The lattice simulation presented is able to establish a link among the energy-landscape theory of protein folding, simulations, and single-molecule folding experiments (Wang and Wolynes, 1995, 1999; Onuchic et al., 1999; Berezhkovskii et al., 1999; Wang, 2003a,b; Lee et al., 2003; Zhou et al., 2003; Gopich et al., 2003a,b; Zhuang et al., 2000, 2002; Jia et al., 1999; Deniz et al., 2000; Schuler et al., 2002; Lipman et al., 2003).

MODEL AND METHODS

The model used for the kinetic simulations has been extensively employed in previous studies (Abkevich et al., 1994; Socci and Onuchic, 1994, 1995). The protein is modeled by a 27-length polymer chain (27-mer) on a three-dimensional cubic lattice. The 27-mer lattice is a renormalized (or reduced) description of small globular proteins. We use a Monte Carlo algorithm with standard polymer local moves—i.e., end, corner flip, and 90° crankshaft (Socci and Onuchic, 1994, 1995). The energy for the heteropolymer is given by

$$E = N_1 E_1 + N_{\mathbf{u}} E_{\mathbf{u}},\tag{1}$$

where E_1 is the nonbonded contact energy between monomers of the same type and E_u is the energy between monomers of different types. N_1 is the number of contacts between monomers of similar type, and N_u is the number of contacts between monomers of dissimilar types. There are only two types of monomers with a hydrophobic or hydrophilic nature. Two limits of the parameters are tested (Chahine et al., 2002):

- 1. High hydrophobicity ($E_1 = -3$ and $E_u = -1$ in arbitrary units, chosen so that typical temperatures are of order one), where biasing toward the folded state is strong ($(E_1 + E_u)/2 = -2$) and the roughness of the landscape is weak (the variance or dispersion is small between native and nonnative contact energies (E_u – E_1) = 2).
- 2. Low hydrophobicity ($E_1 = -3$ and $E_u = +3$), where the biasing toward the folded state is weak ($(E_1 + E_u)/2 = 0$) and the roughness of the landscape is strong (the variance or dispersion is large between native and nonnative contact energies (E_u – E_1) = 6).

We use the previously-designed sequence ABABBBBABBABBABAAABBAAAAAB (Shakhnovich and Gutin, 1993). In this study only one good folding sequence is used. Effects of sequence variations are not analyzed. Monte Carlo sampling with a local move set is used to determine the density of states and to define the kinetics of the model (Abkevich et al., 1994; Socci and Onuchic, 1994, 1995; Chahine et al., 2002). The density of states is determined as a function of energy E, number of native nonbonded contacts Q, and the total number of nonbonded contacts Z. Within the microcanonical ensemble, the free energy of the system can be obtained, and the complete thermodynamics can be determined. Four phases are typically found—i.e., the noncompact unfolded states (so-called random-coil states); compact unfolded states; trapping states; and the native state. The transition temperatures $T_{\rm f}$ (folding) and $T_{\rm g}$ (local trapping) are determined.

We can measure the MFPT to reach the native state for the designed sequence starting from random-coil configurations. By repeating the dynamic Monte Carlo simulations with different initial conditions, one can obtain information about the statistical distributions of these folding times and therefore determine the high-order moments of the FPT. The moments of first-passage time (τ) of events for folding and collapse are generally given by

$$\overline{\tau}^{n} = \int_{0}^{\infty} t^{n} \frac{dP(t)}{dt} dt, \qquad (2)$$

where P(t) is the fraction of unfolded (or uncollapsed) proteins. When the system is above the kinetic minimum temperature (specified in detail in the next section), P(t) exhibits exponential behavior, so $\overline{\tau}^n$ is simply given by

$$\overline{\tau}^{n} = \alpha \frac{\partial^{n}}{\partial \alpha^{n}} \int_{0}^{\infty} e^{-\alpha t} dt = \frac{n!}{\alpha^{n}}.$$
 (3)

The ratio of the n^{th} moment and the mean to the n^{th} power,

$$R_{\rm n} = \overline{\tau^{\rm n}}/\overline{\tau}^{\rm n},\tag{4}$$

can be easily computed in the simulations, and in the high-temperature limit (above the transition temperatures) $R_{\rm n}=n!$. This implies a Poisson distribution, and therefore a single-exponential process. The ratio $R_{\rm n}/n!$ becomes a measure of the deviation from the single-exponential process. As shown below, when the temperature drops, deviations of $R_{\rm n}/n!$ from 1 are observed and one can infer the effects of fluctuations on an FPT. The n-order moment ratio first rises as the temperature drops, then at an even lower temperature, it returns to single-exponential kinetics.

RESULTS

Exponential and nonexponential behavior

Consistent with previous studies (Socci and Onuchic, 1995; Kaya and Chan, 2000, 2002; 2003c), the mean first-passage time (MFPT) as a function of temperature has a U-shaped dependence, with higher values on both the high and low temperature ranges and a kinetic minimum near the folding temperature. We found that above this temperature, the distribution of first-passage times (FPT) is nearly Poissonian, indicating single-exponential kinetics. At the high temperatures, due to the large thermo motions, the different kinetic paths sense roughly similar barriers since the local details are smeared or washed out by the thermo motions. Only the dominant barriers matter. So there are multiple parallel paths, all experiencing similar barriers, resulting in single-exponential kinetics. On the other hand, when the temperature drops below the temperature of the kinetic minimum, the distribution of FPT deviates from Poissonian significantly, indicating nonexponential kinetics and multiple traps. In certain temperature regimes, the distribution at long times approaches a power law. This indicates that the fluctuation in FPT is significant and deviates from the mean. In other words, the FPT is non-self-averaging and the distribution has fatty tails. This indicates the existence of *intermittence*, where rare events can give a significant contribution to the folding event. Due to the ruggedness of the underlying energy landscape at low temperatures, specific discrete paths are themselves different from each other and give distinct contributions to the kinetics. The full distribution of FPT, not only its mean, is needed to characterize the whole system dynamics. At even lower temperatures, the kinetics is back to Poissonian, indicating exponential kinetics and the domination of a single trap. By increasing (or decreasing) hydrophobicity, the distribution deviates less (or more) from Poissonian in the same temperature range.

Folding

The results for the folding FPT moments (second, third, and fourth moments) and their ratios as well as their derivatives with respect to temperature are shown in Fig. 1 (low hydrophobicity) and Fig. 2 (high hydrophobicity). For our sequence, at low hydrophobicity, $T_{\rm g}=1.2$ and $T_{\rm f}=1.6$ (Socci and Onuchic, 1994, 1995). The kinetic minimum of folding $T_0^{\rm fold}$ is near 1.8. For $T>T_0^{\rm fold}$, $R_{\rm n}=n!$; for $T< T_0^{\rm fold}$, $R_{\rm n}/n!>1$; and $R_{\rm n}$ increases significantly. For high hydrophobicity, $T_{\rm g}=1.0$ and $T_{\rm f}=1.3$, the kinetic minimum $T_0^{\rm fold}$ is near 1.5, and one can see a similar behavior: for $T>T_0^{\rm fold}$, $R_{\rm n}=n!$ and for $T< T_0^{\rm fold}$, $R_{\rm n}/n!>1$, but $R_{\rm n}$ does not increase as quickly as for low hydrophobicity.

It is worthwhile to point out that for each of the two models studied, the fastest folding occurs at a model temperature higher than the folding temperature $T_{\rm f}$. As pointed out recently by Chan et al. (2004), this model

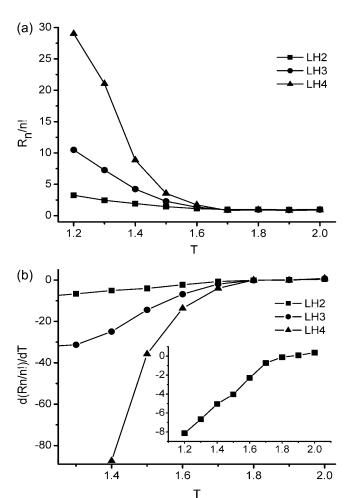
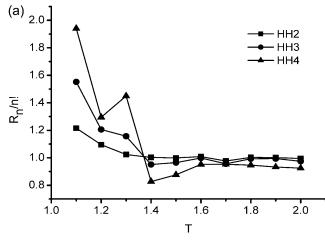


FIGURE 1 Ratios of moments of folding events (a) and their derivatives (b) at low hydrophobicity (*LH*) as function of temperature. The kinetic minimum, at which the $R_n/n!$ start to increase, is at T_0^{fold} . The numbers 2, 3, and 4 refer to R_2 , R_3 , and R_4 , respectively. The smaller plot shows the detail of R_2 .

behavior is not similar to that of many real proteins. It corresponds to a situation in which folding rate would decrease when native stability is increased from the transition midpoint. The underlying reason for this critical shortcoming is because the two-letter models studied here are not sufficiently cooperative, with $T_{\rm f}/T_{\rm g}$ too small to mimic that of real apparent two-state proteins. Although the simulation results presented are useful, these limitations of the model might have significant impact on their relationship with experiments. The cooperativity can play an important role here in solving this discrepancy.

The simulations are in agreement with analytical calculations as well as detailed models (Lee et al., 2003; Zhou et al., 2003) and with other ways of calculating $T_0^{\rm fold}$. This indicates that the ratios of moments or the fluctuations in first-passage times are efficient and sensitive ways to identify the kinetic transition temperatures and the effect of fluctuations. In Figs. 1 and 2 one can see that the derivative of $R_{\rm n}/n!$ provides an effective parameter for locating $T_0^{\rm fold}$. As we can see, the sensitivity on the kinetic fluctuations increases



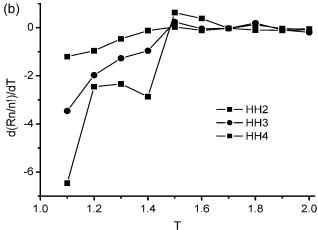


FIGURE 2 Ratios of moments of folding events (a) and its derivatives (b) at high hydrophobicity (HH) as function of temperature. The kinetic minimum, at which the $R_0/n!$ start to increase, is at $T_0^{\text{fold}} = 1.5$. The same notation as in Fig. 1 is used.

rapidly as the order of the moments increases. Although highorder moments will be better indicators than low-order moments, the data requirement is much greater. In principle, there should be a balance in extracting the useful information from the trajectories in the simulations and experiments.

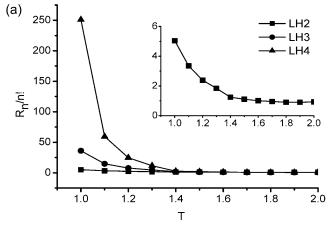
Note that the ratios of the moments will become important at the single-molecule level. In the bulk experiments, the high-order moments are significantly suppressed by the large number of molecules in the sample (on the order of 10⁸ molecules). Furthermore, it is often hard to distinguish the intrinsic dynamics from the observed nonexponential kinetics, since the same data could result from either inhomogeneous distributions of the different protein molecules with exponential kinetics or homogeneous distribution of the intrinsic underlying nonexponential kinetics.

Collapse

Using methods similar to those for the folding simulations, all simulations were repeated to determine the collapse times.

For this study, the collapse time is taken to be at the point when the final state for the collapse is achieved—i.e., when the system reaches the nonbonded contacts Z=25—instead of at the native configuration for the folded state. In the results below we have the same sequence in the same two limits we had before. Now the deviations on R_n are expected to appear when the system reaches the kinetic minimum for collapse $T_0^{\rm col}$. For favorable collapsed states, starting with random-coil conformations, the uncollapsed fraction decays exponentially. However if the glassy trapping regime is reached, the dynamics becomes slow and one observes slow trajectories and nonexponential decays.

For low hydrophobicity, $T_{\rm g}=1.2$, $T_0^{\rm col}=1.6$; and for high hydrophobicity, $T_{\rm g}=1.0$, $T_0^{\rm col}=1.3$. The simulation results of $R_{\rm n}/n!$ and their corresponding derivatives with respect to temperature as a function of temperature are shown in Figs. 3 and 4. Roughly for $T>T_0^{\rm col}$, $R_{\rm n}/n!=1$, and for $T<T_0^{\rm col}$, $R_{\rm n}/n!>1$. Note that $R_{\rm n}$ starts to increase above $T_{\rm g}$ (at T=1.6 for low hydrophobicity, and at T=1.3 for high hydrophobicity), a possible indication of kinetic glassy transition temperature $T_{\rm A}$ postulated in the theories of the short-range random first-



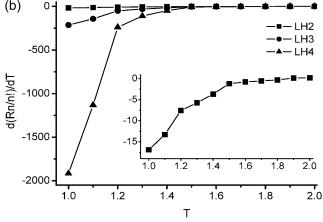


FIGURE 3 Ratios of moments of collapse events (a) and its derivatives (b) at low hydrophobicity (*LH*) as function of temperature. The numbers 2, 3, and 4 refer to R_2 , R_3 , and R_4 , respectively. The kinetic minimum, at which the $R_n/n!$ start to increase, is at $T_0^{\rm col}=1.6$.

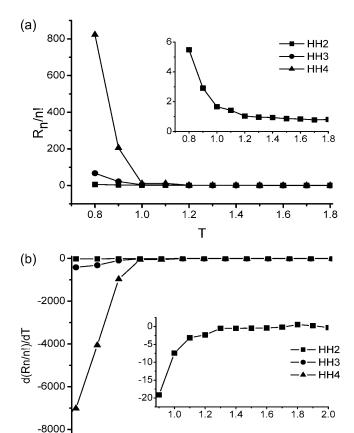


FIGURE 4 Ratios of moments of collapse events (a) and its derivatives (b) at high hydrophobicity (HH) as function of temperature. The kinetic minimum, at which the $R_{\rm n}/n!$ start to increase, is at $T_0^{\rm col}=1.3$. The same notation of numbers as in Fig. 3 is used.

1.4

Т

1.6

1.8

2.0

1.2

1.0

0.8

order phase transitions in glasses (Kirkpatrick et al., 1989). This is an indication that deviation in R_n is a sensitive probe of the dynamics of collapse events (see Figs. 3 b and 4 b for the derivative part).

The increase in R_n below the transition temperatures is hard to quantify. For folding, the increase is more pronounced at low hydrophobicity. For collapse, the behavior is similar at high and low hydrophobicity. The increase in R_n may be related to the relative depth of local traps and the bias toward the native/collapse state, which yields to large fluctuations in the FPT events.

Very low temperature results

The above results on kinetics and fluctuations on folding are valid for temperatures between the folding and the glassy trapping temperatures. At even lower temperature (see Fig. 5), in the case of collapse in the low hydrophobicity limit, R_n drops. At temperature 0.7 we obtain $R_2(=\overline{\tau^2}/\overline{\tau^2})=2.9$, which is quite close to 2! (the high temperature limit of

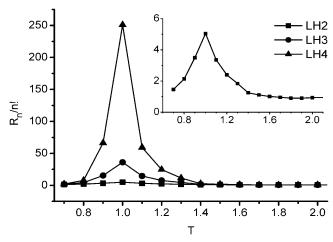


FIGURE 5 Ratios of moments of collapse events at low hydrophobicity (LH) including very low temperatures.

Poisson single-exponential kinetics). The exact numbers are hard to obtain, because at low temperatures a significant percentage of runs do not reach the collapsed state (they reach the Monte Carlo step simulation limit first, 1.0×10^9). Still, the results suggest that the ratios at very low temperature behave much as they do in the high temperature limit, as predicted in analytical calculation ($R_n = \overline{\tau}^n/\overline{\tau}^n = n!$) (Wang, 2004a). Due to the finite size of our systems, kinetics is now controlled by a single barrier.

It is worthwhile to point out that the temperature in the present models cannot be directly identified with the experimental temperature, because the model interactions are temperature-*independent*, whereas real effective intraprotein interactions are temperature-*dependent* (Chan et al., 2004).

In Fig. 6, all kinetic regimes are shown through a plot of the survival population of uncollapsed runs as a function of time. At high temperature (T = 2.0), the system follows an exponential decay. In this regime, multiple diffusive pathways for collapse are available and they are roughly equivalent, yielding a single-exponential characteristic time. At temperatures around the relevant transition temperatures ($T = T_g = 1.0$), fluctuations in τ are at their highest, diffusive paths are not equivalent, and the system exhibits intermittence. One can clearly see that a single exponential does not fit the survival probability. Even a fit to a stretched exponential is not sufficient and the system shows a powerlaw behavior over three decades at times longer than MFPT. At very low temperature (T = 0.7), a single path dominates the process. All three regimes are in agreement with the theoretical predictions (Wang, 2004a).

One can observe two transition temperatures. The high transition temperature corresponds to a transition where the kinetics exhibits changes from the high-temperature exponential to a nonexponential behavior. The low transition temperature (even lower than the glass or trapping transition

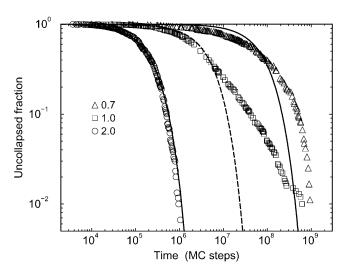


FIGURE 6 Survival probability for collapse (uncollapsed fraction) for the low hydrophobicity case, as in Fig. 5. It shows three different regimes: (\bigcirc) high temperature, T=2.0; (\square) at transition temperature $T=T_{\rm g}=1.0$; and (\triangle) very low temperature, T=0.7. The lines are the exponential fits for each temperature. At high and very low temperature the fits appropriately describe the data ($continuum\ lines$). At the transition temperature one can clearly see the nonexponential behavior (compare with the $dashed\ line$). In the long-time limit between $t=10^6$ and 10^9 , the survival probability is nearly a straight line in the log-log plot implying a power law dependence on time.

temperature) corresponds to the changes in kinetics from higher temperature nonexponential to lower temperature Poisson-exponential.

This kind of kinetic behavior in the fluctuation of the kinetic folding times can be understood physically as follows: At very high temperature, due to the large thermal motion and kinetic energy, the folding has multiple parallel trajectories that cause an average barrier, and therefore the process is a single-exponential one. When the temperature is reduced, the folding event encounters different barriers, and the kinetics becomes multiexponential. When the temperature becomes even lower (below the glassy trapping temperature), the system has only limited states to explore (the others are frozen), and the kinetics is controlled by escapes from a single dominant deep trap. Thus the process becomes a single-exponential one again.

DISCUSSIONS AND CONCLUSIONS

We have studied the fluctuations of protein folding kinetics by examining the ratios of the moments of the first-passage times. This approach can be used to probe the underlying energy landscape structure by kinetics over a broad temperature range. Gruebele's group (Sabelko et al., 1999; Nguyen et al., 2003) observed a "strange" kinetic behavior in which kinetics varies from single-exponential to non-exponential transitions as the temperature decreases. This corresponds to the previous investigation (Kaya and Chan,

2002, 2003c) and our current model prediction in the relatively high temperature regime (the physiological temperature regime), where a change from single-exponential to nonexponential kinetic behavior is observed. Indeed, previous (Kaya and Chan, 2002, 2003c) and current investigations provide evidence that these kinetic features are robust across several different models. This temperaturedependence of the protein dynamics can also be probed in ligand binding experiments. Frauenfelder et al. (1988, 1991) have observed this behavior for the binding kinetics of CO to myoglobin. Sadqi et al. (2003) observed a dynamical slowdown of the hydrophobic collapse upon temperature change. They also observed that, at higher temperatures, as the protein becomes increasingly compact due to a stronger hydrophobic force, there is a slowdown of the dynamics of collapse. This dynamic hydrophobic effect is a hightemperature analog of the dynamic glass transition predicted by theory. In our collapse studies, we also see a similar complex behavior of the kinetics when the temperature changes. To study the role of hydrophobic collapse in folding and its link with the experiments (Sadqi et al., 2003), we plot the evolution of survival probability or the fraction of uncollapse in time. In Fig. 7, we show, under the condition of high hydrophobicity, that the kinetics (the rate) derived from the survival probability at the optimal temperature ($T \sim$ 1.5) is the fastest (measured by the slope of the curve). The kinetics becomes slower when the temperature either increases or decreases beyond the optimal temperature. This is consistent with kinetic slowdown at higher temperatures observed by Sadqi et al. (2003) in the folding experiments (Kaya and Chan, 2000, 2002, 2003c). We also see similar behavior in the low hydrophobicity case in Fig. 8, except that the optimal temperature is at 2.0. In general, the

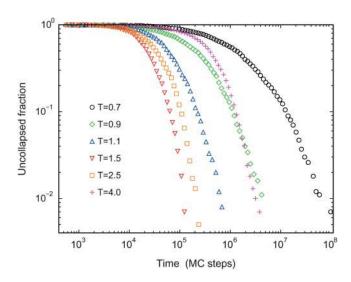


FIGURE 7 Uncollapsed fraction versus time (Monte Carlo steps) for high hydrophobicity case.

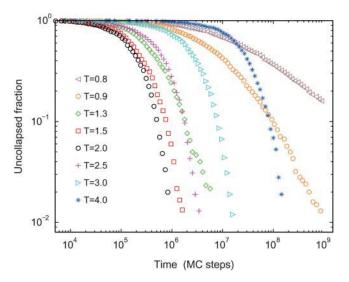


FIGURE 8 Uncollapsed fraction versus time (Monte Carlo steps) for low hydrophobicity case.

average kinetics is slower in the low hydrophobicity case than in the high hydrophobicity case, because of the weaker hydrophobic driving force.

The high-to-low temperature transition from singleexponential to nonexponential kinetics has been also observed in single-molecule experiments. In the work of Xie's group (Yang and Xie, 2002a,b; Yang et al., 2003), reaction dynamics also experiences nonexponential kinetic behavior and the distribution of kinetics follows an approximate power law. The theoretical framework presented here provides a microscopic foundation to interpret these experiments. Other single-molecule experiments also show similar behavior (Zhuang et al., 2000, 2002; Jia et al., 1999; Schuler et al., 2002; Lipman et al., 2003). In future studies, it will be important to explore these kinetic fluctuations upon changing experimental conditions such as temperature and denaturant solvents (which would be equivalent to changing the hydrophobic contents of the system modeled with low and high hydrophobicity in this study).

One of the advantages of this approach is that it provides a link among theory, simulations, and experiments. In the theoretical approach, the first-passage time and its statistical properties can be easily obtained from the simulations following the procedures outlined in this article. In the experiments, information about FPT properties can be obtained from the kinetic folding trajectories (for example, fluorescence). This will stimulate the current round, and the next, of single-molecule experiments and even more detailed simulations, for study of the full range of kinetic behavior.

More simulations at the microscopic level, which will provide the detailed mechanistic features, are also necessary. The only limiting step is that the statistics of the kinetics requires long-time trajectories. Therefore some intermediate

coarse-grained description, for example at the residueresidue level, may be needed to supplement the fully atomistic studies.

This approach proposed for the folding kinetics is quite general. It is not necessarily limited to folding studies. In fact, it is a framework that can be generalized and extended to other systems such as RNA folding, biomolecule binding, viscous liquids, and glassy materials. In fact, such kinetic investigations have already been carried out using an energy landscape framework for viscous liquids and glassy materials (Kirkpatrick et al., 1989; Xia and Wolynes, 2001) and binding of biomolecules (Wang et al., 2004b).

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